A Variant in *PPP4R3A* Protects Against Alzheimer-Related Metabolic Decline

Leigh Christopher, PhD,¹ Valerio Napolioni, PhD,¹ Raiyan R. Khan, BS,¹ Summer S. Han, PhD,² and Michael D. Greicius, MD, MPH,¹ for the Alzheimer's Disease Neuroimaging Initiative

Objectives: A reduction in glucose metabolism in the posterior cingulate cortex (PCC) predicts conversion to Alzheimer's disease (AD) and tracks disease progression, signifying its importance in AD. We aimed to use decline in PCC glucose metabolism as a proxy for the development and progression of AD to discover common genetic variants associated with disease vulnerability.

Methods: We performed a genome-wide association study (GWAS) of decline in PCC fludeoxyglucose F 18 ($[^{18}F]$ FDG) positron emission tomography measured in Alzheimer's Disease Neuroimaging Initiative participants (n = 606). We then performed follow-up analyses to assess the impact of significant single-nucleotide polymorphisms (SNPs) on disease risk and longitudinal cognitive performance in a large independent data set (n = 870). Last, we assessed whether significant SNP influence gene expression using two RNA sequencing data sets (n = 210 and n = 159).

Results: We demonstrate a novel genome-wide significant association between rs2273647-T in the gene, *PPP4R3A*, and reduced [¹⁸F] FDG decline ($p = 4.44 \times 10^{-8}$). In a follow-up analysis using an independent data set, we demonstrate a protective effect of this variant against risk of conversion to MCI or AD (p = 0.038) and against cognitive decline in individuals who develop dementia ($p = 3.41 \times 10^{-15}$). Furthermore, this variant is associated with altered gene expression in peripheral blood and altered *PPPP4R3A* transcript expression in temporal cortex, suggesting a role at the molecular level.

Interpretations: *PPP4R3A* is a gene involved in AD risk and progression. Given the protective effect of this variant, *PPP4R3A* should be further investigated as a gene of interest in neurodegenerative diseases and as a potential target for AD therapies.

ANN NEUROL 2017;82:900-911

Among the many biomarkers for Alzheimer's disease (AD), decline in glucose metabolism in the posterior cingulate cortex (PCC) is one of the earliest, occurring years before symptom onset.¹ Furthermore, metabolic decline in the PCC actually predicts conversion from healthy aging to mild cognitive impairment (MCI), and from MCI to AD,^{2,3} signifying its role in AD progression. The PCC is a central and highly interconnected brain region, with a crucial role in coordinating memory and internally driven cognitive processes.^{4,5} It is also a component of the default mode network, a brain network that is particularly vulnerable in AD.^{6,7} Therefore, molecular pathways involved in declining PCC glucose metabolism may be closely associated with disease progression and worsening of symptoms. Although early decline in PCC glucose metabolism is well established in AD pathology, the genetic contribution to these changes remains unknown. Common genetic variation may influence the extent to which individuals are protected from or predisposed to hypometabolism, and as a result,

View this article online at wileyonlinelibrary.com. DOI: 10.1002/ana.25094

Received May 16, 2017, and in revised form Oct 18, 2017. Accepted for publication Nov 5, 2017.

Address correspondence to Dr Leigh Christopher, Department of Neurology and Neurological Sciences, FIND lab, Stanford University, 780 Welch Road, Stanford, CA. E-mail: lchris@stanford.edu

From the ¹Department of Neurology and Neurological Sciences, FIND Lab, Stanford University, Stanford, CA; and ²Quantitative Sciences Unit, Stanford Center for Biomedical Research (BMIR), Neurosurgery and Medicine, Stanford University, Stanford, CA

Data used in preparation of this article were obtained from the Alzheimer's Disease Neuroimaging Initiative (ADNI) database (adni.loni.usc.edu). As such, the investigators within the ADNI contributed to the design and implementation of ADNI and/or provided data, but did not participate in analysis or writing of this report. A complete listing of ADNI investigators can be found at: http://adni.loni.usc.edu/wp-content/uploads/how_to_apply/ADNI_ Acknowledgement_List.pdf

Additional supporting information can be found in the online version of this article

their risk of developing AD and memory impairment. Genome-wide association studies (GWAS) have identified single-nucleotide polymorphisms (SNPs), or common genetic variants, that are associated with hallmark pathological biomarkers of AD such as beta-amyloid plaques in the brain and phosphorylated tau in the cerebrospinal fluid (CSF).^{8,9} These studies have provided important insight into biological pathways associated with AD; however, the link between PCC metabolic decline and vulnerability to AD is poorly understood. Decline in PCC metabolism correlates well with disease progression. By contrast, amyloid deposition measured with positron emission tomography (PET), appears less sensitive to disease progression.^{10,11} As such, decline in PCC metabolism over time may provide distinct and important insights into biological mechanisms underlying the disease.

The primary aim of this study was to discover SNPs associated with longitudinal decline in PCC glucose metabolism that affect the (1) risk of developing AD and (2) progression of the disease (ie, cognitive decline). Although the pathways linking PCC glucose metabolic decline to AD remain unknown, there is substantial evidence supporting a role for abnormal glucose regulation in AD. Cellular uptake of glucose is closely regulated by oxidative stress signaling pathways.¹² Furthermore, impaired oxidative stress signaling has been shown to lead to reduced glucose metabolism and the eventual development of memory impairment in AD.¹³ Therefore, genes involved in adaptation to oxidative stress or the regulation of glucose metabolism may contribute to PCC metabolic decline and disease vulnerability. Given that the PCC is the first and most pronounced region to undergo metabolic decline, PCC metabolism is a powerful and unexplored endophenotype for investigating unknown genetic contributions to AD. Fludeoxyglucose F 18 ([¹⁸F] FDG) PET measures the uptake of glucose into neurons and astrocytes and can be used as a proxy for neuronal activity. Thus, in order to achieve these aims, we performed a quantitative trait GWAS of longitudinal decline in PCC glucose metabolism, as measured by [18F] FDG PET imaging in the Alzheimer's Disease Neuroimaging Initiative (ADNI) database. Our top candidate from the GWAS was then explored further to determine its effects on AD risk, decline in cognitive performance over time, and gene expression in brain.

Patients and Methods

Participant Details

Participants included in the original imaging GWAS were part of the ADNI (http://www.adni.loni.usc.edu). We restricted the analysis to participants who were white with European ancestry.

TABLE 1. Participant Demographics				
	ADNI	NACC		
Participants (n)	606	870		
APOE*4 (carriers/noncarriers)	265/341	244/626		
Baseline age (years)	74.1 (7.1)	74.9 (8.80.30)		
Education (years)	16.0 (2.8)	16.8 (8.80.94)		
Sex (female/male)	229/377	515/355		
Table demonstrates demographics for each dataset used in the GWAS (ADNI) and follow-up analyses including the analysis for conversion to MCI or AD (NACC) and the analysis for longitudinal cognition (NACC). Values in brackets are standard deviation. ADNI = Alzheimer's Disease Neuroimaging Initiative; NACC = National Alzheimer's Coordinating Centre.				

Individuals included in the imaging GWAS had [18F] FDG PET data available at two time points at least a year apart. This included a total of 606 ADNI participants (mean age of 74 years \pm 7.1 [standard deviation] mean education of 16 years \pm 2.8, 43.7% APOE*4 carriers, and 37.8% female; Table 1). Participants for the analysis of conversion to MCI or AD and of longitudinal cognitive change were from the National Alzheimer's Coordinating Centre (NACC).14-16 This is a data set including healthy elderly and AD participants with longitudinal clinical and cognitive data (Table 2). Participants for the brain RNA sequencing (RNA-Seq) analysis were part of the MAYO RNA-Sequencing study¹⁷ and the Rush Memory and Aging Project (MAP)¹⁸/Religious Orders Study (ROS).¹⁹ These are data sets with RNA-Seq transcript data and genotype data for healthy and AD participants (Table 2). For the analysis of conversion to MCI or AD, we included healthy participants at baseline from NACC who had longitudinal clinical data and genotype data available (n = 870) in order to track their conversion over time (mean age of 74.9 years ± 8.8, mean education of 16 years \pm 8.8, 28% APOE*4 carriers, and 59.2% female). For the longitudinal cognitive change analysis, we included all NACC participants who had longitudinal cognitive data at two time points at least a year apart and available genotype data (n = 851). All of the participants for this analysis overlapped with the participants included in the conversion analysis. For the RNA-Seq analysis, we used two data sets from ROS/MAP and MAYO which provide RNA-Seq data from the dorsolateral prefrontal cortex and temporal cortex, respectively. For ROS/MAP, we included all participants with available dorsolateral prefrontal cortex (DLPFC) RNA-Seq data and genotype data (n = 220, mean age of death 84.4 ± 4.5 , mean education of 16.7 ± 3.4 , 25.9% APOE*4 carriers, and 66.4% female). For the MAYO data set, we included all participants with available temporal cortex RNA-Seq data and genotype data (n = 159, mean age of death 80.3 ± 7.8 , 32.9% APOE*4 carriers, and 55% female). All participants provided written informed consent, and the protocols were approved by their

TABLE 2. RNA-Seq Participant Demographics			
	ROS/MAP RNA-Seq	MAYO RNA-Seq	
Participants (n)	220	149	
APOE*4 (carriers/noncarriers)	57/163	49/100	
Age at death (years)	84.4 (4.5)	80.3 (7.8)	
Sex (female/male)	146/74	82/67	
Disease group (healthy/MCI/AD)	68/63/89	72/0/77	
Postmortem interval (years)	7.0 (4.6)	6.6 (6.3)	
Education (years)	16.6 (3.4)	_	
Tissue type	DLPFC	Temporal cortex	
Table demonstrates demographic information for each data set used in the RNA-Seq analysis. Values in brackets are standard deviation. MCI = mild cognitive impairment; AD = Alzheimer's disease; DPLFC = dorsolateral prefrontal cortex; BA = Broadmann area.			

respective review boards. NACC, ROS/MAP, and MAYO study data were retrieved from the "NIA Genetics of Alzheimer's Disease Data Storage Site" (NIAGADS) and the Sage Bionetwork's Synapse project. The ADNI was launched in 2003 as a publicprivate partnership, led by Principal Investigator Michael W. Weiner, MD. The primary goal of ADNI has been to test whether serial magnetic resonance imaging, PET, other biological markers, and clinical and neuropsychological assessment can be combined to measure the progression of MCI and early AD. This was a reanalysis of de-identified data available from shared data repositories. Accession codes for the publicly available data used for the present study are reported in the Data Software and Availability section. The study protocol was granted an exemption by the Stanford Institutional Review Board because the analyses were carried out on "de-identified, off-the-shelf" data.

Method Details

IMAGING ANALYSIS. All participants from ADNI-1 and ADNI-GO/2 with longitudinal [18F] FDG-PET available were included in the GWAS. [18F] FDG is used to measure resting cerebral metabolic rate of glucose uptake in the brain and is an early marker of neurodegeneration. [18F] FDG-PET scans were acquired and preprocessed using a region of interest (ROI) approach as described previously.²⁰ In brief, ROIs were created using a meta-analytic approach to determine regions most frequently associated with glucose metabolic changes and AD. The PCC was selected as the primary ROI for this study. In previous work by Jagust et al, the most thoroughly preprocessed version of the ADNI [18F] FDG-PET data were downloaded and uptake in each ROI was normalized to uptake in the pons as a reference region, resulting in summary values of [18F] FDG-PET uptake in a number of AD-related brain regions.²⁰ These ROI data were used here to calculate decline in PCC [¹⁸F] FDG as the mean annual percent decline from baseline between the first and last [18F] FDG PET scan (Fig 1A). Only participants who had at least 1 year of longitudinal follow-up were included.

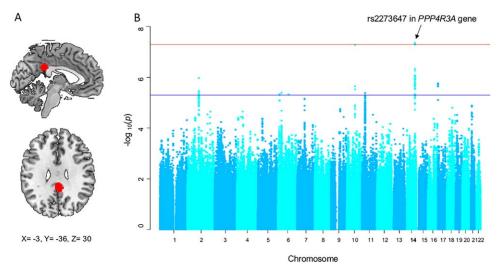


FIGURE 1: A common variant in *PPP4R3A* is associated with less decline in posterior cingulate glucose metabolism. (A) PCC ROI overlaid on a template brain in MNI space illustrating the region where [18 F] FDG PET decline was measured for each participant. MNI coordinates are displayed below. (B) Manhattan plot demonstrating the results of the GWAS including all ADNI participants. The blue line represents a suggestive association threshold (1×10^{-5}), and the red line indicates the genome-wide association threshold (5×10^{-8}). Arrow indicates significant SNP passing genome-wide significance threshold located on chromosome 14 in an intronic region of the *PPP4R3A* gene. ADNI = Alzheimer's Disease Neuroimaging Initiative; [18 F] FDG = fludeoxyglucose F 18; GWAS = genome-wide association studies; MNI = Montreal Neurological Institute; PCC = posterior cingulate cortex; PET = positron emission tomography; ROI = region of interest; SNP = single-nucleotide polymorphism.

Participants in NACC were used as an independent data set to examine, respectively, the effect of significant SNPs obtained from the GWAS in ADNI on (1) risk of conversion to MCI or AD as assessed by the clinical diagnosis at each visit and (2) disease progression (ie, longitudinal cognitive performance). RNA-Seq data from DLPFC were also available for participants in the ROS and MAP studies and in participants from the MAYO study, and these were used to test the functional impact of significant SNPs on brain transcript expression levels.

Genotyping and Imputation

ADNI GWAS data were downloaded from the ADNI database. Genotyping in ADNI was performed using blood DNA samples with three genotyping arrays: Illumina 610-Quad, Illumina-OmniExpress, or Illumina HumanOmni2.5-4v1 as previously described.²¹ Genotype data underwent standard quality control including identity checks, cryptic relatedness (identity-by-descent > 0.0625), sample exclusion for call rate <95%, SNP exclusion for call rate <95%, Hardy-Weinberg equilibrium (HWE) of $p < 1 \times 10^{-6}$, and minor allele frequency (MAF) < 1%. Unlinked SNPs (linkage disequilibrium [LD] pruning at $r^2 = 0.1$) passing the quality control criteria were used to determine the first 10 principal components for population structure using EIGENSTRAT²² and to remove population structure outliers. Imputation was performed using IMPUTE2 software, using the European population from the 1000 Genomes Project, Phase 3 as a reference panel.²³ After imputation, we excluded SNPs with an info quality score <0.9, MAF < 0.05, and HWE ($p < 5 \times 10^{-6}$). This yielded a total of 2,820,932 SNPs for the GWAS analysis. A subset of ADNI subjects (n = 818) was also whole-genome sequenced, and available data were used to confirm the results obtained from SNP imputation. VCF files were annotated using SNPeff²⁴ and analyzed by PLINK 1.9.25 GWAS data were also used for obtaining the genotype information for significant SNP(s) in our follow-up analyses. These data sets also underwent the same quality-control pipeline and imputation described above.

Quantification and Statistical Analysis

GWAS. The GWAS was performed using PLINK software. The analysis was run using a linear regression under the assumption of an additive genetic model. Covariates included baseline age, sex, education, disease status, and the first three principal components for population structure. We did not include APOE*4 as a covariate in the initial GWAS, because of the colinearity between disease status and APOE*4 dosage. The suggestive association threshold was $p < 1 \times 10^{-5}$, and the threshold for genome-wide significance was $p < 5 \times 10^{-8}$. This is a more conservative, consensus threshold for genome-wide significance based on correction for 1 million independent tests.²⁶ We aimed to discover SNPs passing the genome-wide significant threshold. Manhattan plots and QQ plots were generated using the package "qqman" in R (version 3.3.1; R Foundation for Statistical Computing, Vienna, Austria). Regional association plot was created using LocusZoom.

COMPETING RISKS REGRESSION ANALYSIS. We ran a competing risks regression analysis developed by Fine and Gray to evaluate the risk of conversion to MCI or AD by genotype (of significant SNPs), while accounting for death as a competing risk.²⁷ This analysis was carried out using the "cmprsk" package in R.28 Competing risks analysis is a type of time-toevent analysis that aims to accurately estimate the marginal probability of an event in the presence of competing events such as death. This approach is relevant in this study because the participants are elderly and death may occur before the event of interest (ie, conversion to MCI or AD) is observed, which can produce bias in risk estimates. We included all participants from the NACC data set who were healthy at baseline, had genotyping data available, and had at least 1 year of follow-up to assess the effect of significant SNPs on risk of conversion. Age at entry, APOE*4 status, sex, and education were included as covariates in the analysis (n = 870; Table 1). Disease was not included as a covariate because all participants were healthy at baseline.

LINEAR MIXED-EFFECTS ANALYSIS OF LONGITUDINAL COGNITION. We tested the association between significant SNPs and longitudinal change in cognitive performance using participants from the NACC data set who had longitudinal neuropsychological testing data available (n = 851). We used the Logical Memory Delayed Recall Score (out of 25) to assess episodic memory, the Boston Naming Task (BNT; out of 30) to assess language, and the Mini Mental State Examination (MMSE; out of 30) to assess global cognitive function. These tests are known to be sensitive measures of cognitive decline in AD and were acquired longitudinally in the NACC data set. We tested all participants (n = 851) who were healthy at baseline who had available genotype information. We analyzed the data using a linear mixed-effects model implemented in R, testing for the association between genotype and the change in cognitive performance over time, that is, a time by genotype interaction. This model accounts for correlations among repeated measurements within each participant, and permits varying observation periods and varying rates of decline among individuals. In addition to genotype, the time of measurement, and the interaction between these two variables, we included age at baseline, APOE*4 dosage, final diagnosis, education, and sex as covariates.

RNA-SEQ ANALYSIS. We tested whether transcript expression was altered between AD cases and controls for genes containing significant SNPs from the GWAS. We then tested the effect of significant SNPs on transcript expression of genes of interest using the ROS/MAP and MAYO RNA-Seq data sets from DLPFC tissue and temporal cortex tissue, respectively. We used the general linear model implemented in R to perform a regression analysis looking at the effect of genotype on transcript expression for genes containing significant hits in the GWAS. All analyses included age at death, *APOE**4 status, sex, disease status, postmortem interval (PMI), and education (not available in the MAYO data set) as covariates. We removed outliers greater than 2 absolute standard deviations from the mean to

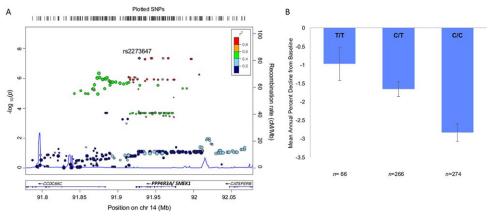


FIGURE 2: Regional association and pattern of PCC [18 F] FDG decline for rs2273647 in *PPP4R3A*. (A) Regional association plot demonstrating regional linkage disequilibrium for rs2273647 with other SNPs. (B) Longitudinal decline in PCC glucose metabolism by rs2273647 genotype. The T allele is associated with significantly less decline in glucose metabolism. [18 F] FDG = fludeoxyglucose F 18; PCC = posterior cingulate cortex; SNPs = single-nucleotide polymorphisms.

exclude the possibility of sampling errors. We also examined two large publicly available online peripheral blood gene expression data sets to test whether significant SNPs modified gene expression in blood (https://eqtl.onderzoek.io/index.php?page=info & http://genenetwork.nl/bloodeqtlbrowser/).

Data and Software Availability

GWAS data and [¹⁸F] FDG PET imaging data used in the present study were retrieved through ADNI (http://adni.loni.usc. edu). GWAS data sets for follow-up competing risks regression and linear mixed-effects analyses were retrieved through NIAG-ADS (https://www.niagads.org) [ADC1 (NG00022), ADC2 (NG00023), ADC3 (NG00024)] for the NACC participants and [ROSMAP (NG00029)] for the ROS/MAP participants. Clinical and cognitive visit data for the NACC participants were obtained by request from the longitudinal uniform data set (RDD-UDS). RNA-Seq data in ROS/MAP participants and MAYO participants were obtained from Sage Bionetwork's Synapse project (https://www.synapse.org, syn3388564).

Results

GWAS Identifies a Variant in PPP4R3A Associated With Decline in PCC [¹⁸F] FDG PET

There was a total of 606 ADNI participants included in the GWAS for PCC decline. Participants included a mixture of 192 healthy adults, 335 MCI patients, and 79 AD patients.

We identified 130 SNPs that passed the suggestive association threshold (Fig 1B; Supplementary Table 1). These SNPs corresponded to nine loci on seven separate chromosomes. Only one SNP passed the genome-wide significance threshold (5×10^{-8}). This was rs2273647, an intronic variant located in the gene, *PPP4R3A* (OMIM*610351, on chr. 14q32.12; Fig 1B; MAF = 0.326; $\beta = 1.303$; $p = 4.44 \times 10^{-8}$; Fig 2A; Supplementary Table 1). The SNP, rs2273647, is located in a transcription factor

binding site near a coding region of the gene *PPP4R3A* (alternate name *SMEK1*). One other SNP came close to genome-wide significance located on chromosome 10 in an intronic region of the gene *CDH23* (rs754726; $\beta = -2.98$; $p = 5.29 \times 10^{-8}$; see Supplementary Table 1). Rs2273647 in *PPP4R3A* was in high LD (r² > 0.8) with other SNPs located nearby on chromosome 14 (Fig 2B). We generated a Q-Q plot to compare the observed versus the expected *p* values for the GWAS. The genomic inflation factor λ was equal to 1.00, indicating that there is no systemic inflation of the observed *p* values in our data set.

The minor allele of rs2273647-T demonstrated a protective effect on PCC glucose metabolic decline, with the homozygous minor allele group showing the least reduction in PCC glucose metabolism followed by the heterozygous group, and the homozygous wild-type groups (Fig 2B). We determined that rs2273647 was an imputed SNP. Therefore, we checked the concordance rate between rs2273647 genotype in individuals who had whole-genome sequencing data in ADNI. There was a 98% concordance rate between rs2273647 genotype and whole-genome sequencing data in ADNI, confirming the validity of imputation for rs2273647.

There were 274 individuals who were homozygous wild type (C/C), 266 who were heterozygous (C/T), and 66 who were homozygous recessive (T/T) at rs2273647 (Table 3). There were no significant differences in baseline age, education, sex, or disease status between the three genotypes groups. There was a significant difference in the proportion of *APOE**4 carriers versus noncarriers (p = 0.011) across rs2273647 genotype groups. When the GWAS was rerun to include *APOE**4 status as a covariate in addition to the original covariates, the effect of rs2273647-T remained quite strong ($p = 1.67 \times 10^{-7}$).

TABLE 3. ADNI Participant Demographics According to Disease Status				
	Healthy Controls (n = 192)	MCI (n = 335)	AD (n = 79)	P
APOE*4 (carriers/noncarriers)	46/146	161/174	56/23	$4.6 \times 10^{-13*}$
Baseline age (years)	75.1 (5.2)	72.9 (7.7)	76.6 (7.2)	0.3
Education (years)	16.3 (2.7)	16.1(2.7)	14.9 (3.2)	0.9
Sex (female/male)	78/114	115/220	29/50	0.35
PPP4R3A rs2273647 genotype (C/C, C/T, T/T)	91/81/20	143/151/41	40/30/9	0.67

This table includes demographic information for individuals from ADNI included in the imaging GWAS. Values in brackets are standard deviation.

p < 0.05 significant difference in frequencies between groups with Pearson's chi-squared test.

ADNI = Alzheimer's Disease Neuroimaging Initiative; MCI = mild cognitive impairment; AD = Alzheimer's disease.

PPP4R3A rs2273647-T Affects Risk of Conversion to MCI or AD

We aimed to assess whether the significant SNP was associated with risk of clinical conversion from healthy aging to MCI or AD. In order to evaluate the risk by genotype, we performed a competing risks regression analysis with death as the competing risk. Participants from the NACC data set who were healthy controls at baseline had longitudinal clinical visit data and had available genotype information were included in the analysis (Table 1, Table 4; n = 870). These subjects are independent from those used in the ADNI GWAS. For the 870 participants included in the analysis, the mean duration of follow-up was 4.3 years and the maximal follow-up time was 7 years. Risk of conversion was considered the risk of conversion from healthy to MCI or from healthy directly to AD (whichever conversion came first). Genotype was coded using an additive model (by dosage of the minor allele). Genotype for rs2273647 was

significantly associated with risk of conversion to MCI or AD in 870 healthy controls. The minor allele (T) dosage was significantly associated with a reduced probability of developing MCI or AD (Fig 3A; hazard ratio [HR] = 0.76; p = 0.038; n = 870). Although the difference in risk between the heterozygote and homozygote T allele carriers was small, there was a more dramatic difference in the risk of conversion between the carriers and noncarriers of the T-allele characteristic of a dominant effect. Therefore, we also ran the competing risks regression using a dominant model (ie, minor allele carriers [CT and TT] vs noncarriers [CC]). There was a significant dominant effect on risk of conversion (Fig 3B; HR = 0.70; p = 0.041; n = 870), with T-allele carriers demonstrating a significantly lower risk compared to noncarriers.

To reduce the risk of bias attributed to the inclusion of individuals with a very short duration of followup, we restricted the analysis to those participants who

TABLE 4. Demographics for NACC Participants (Healthy at Baseline) According to PPP4R3A rs2273647 Genotype				
	T/T (n = 130)	C/T (n = 381)	C/C (n = 359)	p
APOE*4 (carriers/noncarriers)	36/94	111/270	97/262	0.81
Baseline age (years)	74.6 (8.8)	74.9 (8.8)	75.1(8.7)	0.54
Education (years)	16.6 (7.8)	16.8(8.9)	16.8 (9.2)	0.81
Sex (female/male)	68/62	237/144	210/149	0.47

deviation. NACC = National Alzheimer's Coordinating Centre.

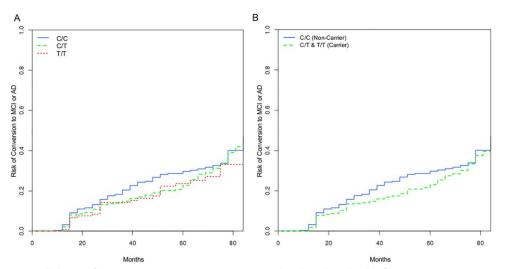


FIGURE 3: The minor allele (T) of rs2273647 in *PPP4R3A* is associated with reduced risk of conversion to MCI or AD. (A) Cumulative incidence functions demonstrating the risk of conversion to MCI or AD for three genotypes of rs2273647 over a 7-year time period using an additive model. T allele dosage was significantly associated with the risk of conversion to MCI or AD over time after controlling for *APOE**4 status, age at entry, sex and education (HR = 0.76; p = 0.038; n = 870). (B). Cumulative incidence functions demonstrating the risk of conversion to MCI or AD using a dominant model. T-allele carriers demonstrate significantly lower risk of conversion to MCI or AD over time after controlling for *APOE**4 status, age at entry, sex and education (HR = 0.70; p = 0.041; n = 870). AD = Alzheimer's disease; HR = hazard ratio; MCI = mild cognitive impairment.

had a minimum of 2 and 3 years of follow-up in separate analyses and tested both the additive and dominant models. When we restricted the sample to those with a minimum of 2 years of follow-up (n = 735; meanfollow-up time of 4.8 years), there was a significant additive effect of genotype on risk of conversion (HR = 0.72; p = 0.01; n = 735) and an even stronger dominant effect on risk of conversion (HR = 0.58; p = 0.0014; n = 735). When we restricted the sample to those with a minimum of 3 years of follow-up (n = 635; mean follow-up time of 5.2 years), genotype was no longer significantly associated with risk of conversion for the additive model (p = 0.13); however, there was a strong significant association between genotype and risk of conversion for the dominant model (HR = 0.63; p = 0.006; n = 635). These results suggest that genotype has a consistent and significant impact on the risk of developing disease. When comparing models, the dominant model provided a better model fit with lower Bayesian information criterion. Therefore, we used a dominant model as the default for the remainder of the follow-up analyses and reported additive model results as well.

PPP4R3A rs2273647-T Affects Longitudinal Cognitive Performance

Next, we aimed to assess the relationship between the *PPP4R3A* variant and cognitive performance over time. We performed a linear mixed-effects analysis on all NACC participants to assess the association between rs2273647-T and the change in cognitive performance (memory, language and global cognition) over time, by

evaluating time by rs2273647 interaction. All participants who were healthy at baseline had longitudinal cognitive data, and genotype data available were included in the model and classified according to final diagnosis (n = 819 for BNT, n = 822 for MMSE, and n = 854 forLogical Memory Delayed Recall). Our analysis showed that cognitive decline was modified by the genotype of rs2273647, with significant interaction for time by genotype observed on both BNT ($p = 1.37 \times 10^{-6}$) and MMSE ($p = 3.41 \times 10^{-15}$) in individuals with a final diagnosis of AD (Fig 4A,B); the extent of cognitive decline over time was reduced among individuals with the T allele in rs2273647 versus wild-type homozygotes, demonstrating its protective effect on cognitive performance. The results were also significant when using the additive model, for both the time by genotype effect on BNT (p < 0.05) and MMSE $(p < 5 \times 10^{-6})$. All effects were significant after controlling for age at baseline, APOE*4 dosage, final diagnosis, education, and sex as covariates.

There were no significant time by genotype interactions in individuals with a final diagnosis of healthy or MCI, whose cognitive performance remained relatively stable over time (Fig 4A,B). There were no significant interactions in any of the three groups on longitudinal logical memory (Fig 4C).

PPP4R3A rs2273647-T Affects Gene Expression

Last, we wanted to assess whether *PPP4R3A* transcript expression is altered in disease and whether rs2273647-T has a functional impact on gene expression. First, we identified the most highly expressed transcripts in brain

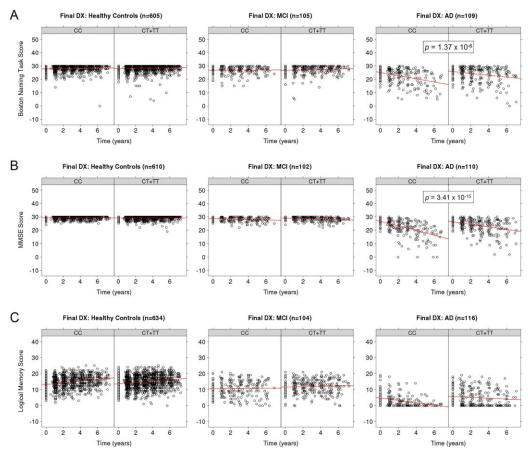


FIGURE 4: Cognitive decline is significantly modified by genotype in individuals with a final diagnosis of AD. (A) Participants who develop AD demonstrate a significant time by genotype interaction on BNT scores ($p = 1.37 \times 10^{-6}$; n = 819), indicating a protective effect on language decline. (B) Participants who develop AD demonstrate a significant time by genotype interaction on MMSE scores ($p = 3.41 \times 10^{-15}$; n = 822), indicating a protective effect on global cognitive decline. Interactions were significant after controlling for baseline age, education, *APOE**4 dosage, and sex. (C) Time by genotype effect was not significant for longitudinal memory scores in any diagnostic category. AD = Alzheimer's disease; BNT = Boston Naming Task; Final DX = final diagnosis; MCI = mild cognitive impairment; MMSE = Mini Mental State Examination.

tissue for PPP4R3A. There were two transcripts that were fully annotated, protein coding and expressed in brain: ENST00000554684 and ENST00000555462. The mean expression in FPKM (fragments per kilobase transcript per million mapped of reads) of ENST00000554684 in DLPFC and temporal cortex tissue was 1.41 (± 0.89) and 28.59 (± 5.34), respectively. The mean expression in FPKM of ENST00000555462 in DLPFC and temporal cortex was 1.39 (±0.65) and 4.77 (± 0.98), respectively. First, we aimed to test whether there were differences in the expression of PPP4R3A between AD cases and healthy controls in the RNA-Seq data sets from DLPFC and temporal cortex (Table 2, Table 5). We found a significant difference between cases and controls in ENST00000554684 expression after controlling for age at death, sex, APOE*4 status, and PMI. The healthy controls demonstrated lower expression compared to AD (Fig 5B; p = 0.0038; n = 159), suggesting that transcript expression of this gene is altered by disease. There were no significant differences between

AD and healthy controls within DLPFC tissue for either transcript.

Next, we aimed to assess whether rs2273647 genotype is associated with altered transcript expression. The rs2273647-T genotype was associated with lower *PPP4R3A* ENST00000555462 transcript expression in temporal cortex of healthy control participants from the MAYO data set, with carriers of the minor allele demonstrating lower transcript expression compared to noncarriers (Fig 5A; p = 0.011; n = 159). Our results were also significant when using an additive model (p = 0.015). There was no significant difference in expression of this isoform in AD participants. Furthermore, there was no effect of genotype on transcript expression in the DLPFC tissue from the ROS/MAP data set.

Finally, we took advantage of two, large, web-based expression quantitative trait loci (eQTL) datasets to test the effect of rs2273647-T dose on *PPP4R3A* expression in peripheral blood. Results in these data sets are only available for the additive model. We found that

TABLE 5. Demographics for Mayo RNA-Seq Data From Temporal Cortex Stratified by Disease			
	AD (n = 77)	Healthy Controls (n = 72)	
APOE*4 (carriers/noncarriers)	40/37	9/63*	
Age at death (years)	80.2 (7.5)	80.3 (8.2)	
Sex (female/male)	47/30	35/37	
There were no significant differences between groups in any of the variables other than <i>APOF</i> *4 status. Healthy controls had a signifi-			

variables other than APOE*4 status. Healthy controls had a significantly different proportion of APOE*4 carriers than AD cases (* $p = 7.5 \times 10^{-7}$, Pearson's chi-squared test). Values with brackets are standard deviation. RNA-Seq = RNA sequencing; AD = Alzheimer's disease.

rs2273647 was an eQTL, lowering the expression of

PPP4R3A in blood with increasing dosage of the minor allele in a large meta-analysis of nine data sets including 5,311 samples $(p = 4.75 \times 10^{-10}; Z = -6.23)$.²⁹ We were able to confirm this result in a second blood eQTL data set of 4,896 participants of European ancestry $(p = 2.53 \times 10^{-10};$ https://eqtl.onderzoek.io), providing further support for the involvement of this variant in modifying gene expression in blood.³⁰

Discussion

Previous GWAS studies using imaging markers including hippocampal atrophy, beta-amyloid, and CSF tau have

provided insight into the mechanisms underlying genetic vulnerability to AD, implicating a number of genes associated with these AD endophenotypes.^{8,9,21,31} The significant advantage of using PCC metabolic decline as an endophenotype in the present study is that it is a consistent early biomarker for AD that has been shown to correlate well with disease progression, to a greater extent than beta-amyloid deposition.^{10,11} Thus, using it as an endophenotype has the potential to reveal important genes that are closely associated with the mitigation (or exacerbation) of disease progression.

In this study, we demonstrate that rs2273647-T is associated with less metabolic decline in the PCC, a reduced risk of conversion to MCI or AD, and reduced cognitive decline. Previous studies have tested for associations between known AD risk variants and imaging markers of AD progression, including PCC hypometabolism³²; however, this is the first study to identify a novel genome-wide association between a common variant and longitudinal PCC metabolic decline.

Taken together, these results suggest that the T allele confers a protective advantage by preserving brain glucose uptake in the face of AD pathology. We show that rs2273647-T is associated with a reduced risk of conversion to MCI or AD, suggesting that PPP4R3A plays a role in predisposition to disease. We also show that rs2273647-T is protective against language and global cognitive decline, in those individuals who progress to AD over their follow-up period. The lack of significant interaction effects in those who remain healthy or convert to MCI in our study is likely attributed to the

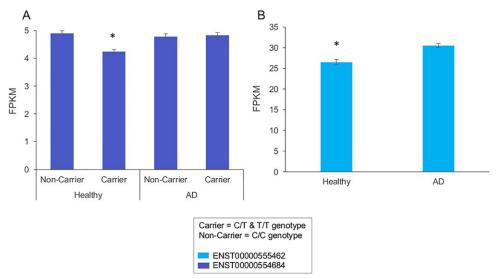


FIGURE 5: Effect of genotype and disease status on PPP4R3A transcript expression. (A) There is a significant dominant effect of genotype on transcript expression (ENST00000555462) in the temporal cortex of healthy controls, with T-allele carriers demonstrating lower expression compared to noncarriers (p = 0.011; n = 159). There is no significant effect of genotype on transcript expression in AD. (B) PPP4R3A transcript expression (ENST00000554684) is significantly lower in healthy control compared to Alzheimer's disease patients in temporal cortex (p = 0.0038; n = 149). AD = Alzheimer's disease; FPKM = fragments per kilobase of transcript per million mapped reads.

fact that many of these participants remained healthy for a large proportion of the study and do not show substantial decline regardless of genotype (as is apparent in Fig 4), thus obscuring the effect of rs2273647-T. Our findings demonstrate a strong protective effect in those individuals who convert to AD, especially against global cognitive decline, emphasizing that individuals carrying rs2273647-T perform better despite the onset of disease.

In order to provide a functional link between PPP4R3A at the molecular level and the clinical level, we aimed to assess whether rs2273647 genotype was significantly associated with PPP4R3A transcript expression. We were able to detect a significant effect of genotype in healthy controls for one of the protein-coding transcripts in temporal cortex, whereby healthy control carriers demonstrated lower transcript expression. We also demonstrated that transcript expression is significantly lower in healthy controls compared to AD, providing further evidence that PPP4R3A is altered in disease. The reason for healthy control carriers demonstrating lower transcript expression, but not AD participants, is unclear; however, it is possible that the effect of disease on transcript expression outweighs the effect of the protective genotype. The lack of replication of these results in the DLPFC may be attributed to a regional effect of rs2276347 on transcript expression within the temporal cortex, which is more strongly affected by AD progression. Gene expression changes appear to be triggered by the onset of AD-associated pathology, which is present earlier and to a greater extent in the temporal cortex than in the frontal cortex. Further work will be needed to determine the regionally specific effects of rs2273647 on gene expression in the brain.

In previous work, PPP4R3A was shown to be involved in gluconeogenesis. Furthermore, reduced expression of PPP4R3A was directly linked to lower fasting blood glucose levels, whereas increased PPP4R3A expression was linked to insulin resistance.33 This strongly suggests that PPP4R3A plays an important role in influencing glucose uptake in humans, and that increased PPP4R3A expression may be detrimental. More tellingly, the SNP identified here has been associated with reduced fasting blood glucose levels ($\beta = -0.015$; $p = 1.7 \times 10^{-4}$).³⁴ Furthermore, we confirmed that it is an eQTL in blood in two independent data sets, significantly reducing gene expression in T-allele carriers.^{29,30} Taking this into consideration with our findings, rs2273647-T carriers may have reduced gene expression and reduced fasting blood glucose levels, which could contribute directly to a protective effect on [18F] FDG decline. Conversely, higher expression of PPP4R3A may result in insulin resistance and reduced brain glucose uptake, which has been linked to AD.^{12,35} Another

possibility is that T-allele carriers experience a lower risk for diabetes as a result of their differential glucose regulation, thus indirectly affecting AD risk and brain metabolic decline.³⁶ Reduced brain glucose uptake is an early marker of neurodegeneration, and not necessarily specific to AD. Therefore, it is possible that PPP4R3A may influence vulnerability to multiple neurodegenerative diseases.37 Although the exact mechanism of action of PPP4R3A in AD is unknown, abnormal regulation of insulin signaling and glucose metabolism is associated with greater oxidative stress, and thus PPP4R3A may play a role in the predisposition of neuronal cells to oxidative damage and metabolic dysfunction associated with neurodegenerative disease pathology. Interestingly, smk-1 (PPP4R3A ortholog) has been shown to play a direct role in mediating longevity through the insulin-signaling pathway.^{38,39} Furthermore, accumulating evidence suggests that the effective regulation of insulin signaling promotes healthy aging and is protective against toxic age-related protein aggregation, including amyloid-beta.40-42 Thus, alterations in PPP4R3A may help slow the onset and accumulation of AD pathology through the modification of insulin-signaling pathways. In a follow-up search for previous associations between PPP4R3A and AD risk, we found that a distinct variant in PPP4R3A passed the suggestive threshold for association with risk of AD in a familybased GWAS, lending additional support for the involvement of this gene in AD.43 Further investigations will be needed to identify the protective mechanisms by which PPP4R3A affects disease vulnerability.

One limitation to our study is that we have not been able to determine definitively whether the true biological effect of rs2273647-T is dominant or additive. As is conventionally done in GWAS studies, we identified this variant in our initial analysis with an additive model. We observed in the follow-up analyses, however, that the effect of genotype on disease risk, progression, and gene expression was stronger when rs2273647 genotype was classified according to a dominant model. We therefore returned to the original GWAS and checked the p value of our SNP assuming a dominant model and found that it remains quite significant at $p = 1.32 \times 10^{-7}$. On balance, our results support an additive model with the dominant model appearing stronger mainly in those analyses (such as gene expression in brain) with smaller sample sizes and relatively few homozygous recessive subjects.

In conclusion, we identify a novel protective variant in the gene *PPP4R3A* associated with reduced glucose metabolic decline. We were able to replicate the protective effect in an independent data set, demonstrating that the minor allele at rs2273647 is associated with a reduced risk of developing AD and a slower rate of cognitive decline in subjects who ultimately develop AD. Importantly, our findings strongly support a role for *PPP4R3A* in AD vulnerability and progression. Furthermore, this variant affects *PPP4R3A* gene expression, indicating a functional effect. Although the specific biological pathways underlying the role of *PPP4R3A* in AD vulnerability require further investigation, the results reported here suggest that *PPP4R3A* should be considered as a potential therapeutic target in AD.

Acknowledgment

Supported by the CIHR Fellowship Award, NIH:P50 AG047366, McKnight Memory and Cognitive Disorders Award, Feldman Foundation CA, The JNA Foundation, The J. W. Bagley Foundation, and The Mervin and Roslyn Morris Educational and Philanthropic Fund.

Data collection and sharing for this project was funded by the Alzheimer's Disease Neuroimaging Initiative (ADNI) (National Institutes of Health Grant U01 AG024904) and DOD ADNI (Department of Defense award number W81XWH-12-2-0012). ADNI is funded by the National Institute on Aging, the National Institute of Biomedical Imaging and Bioengineering, and through generous contributions from the following: AbbVie, Alzheimer's Association; Alzheimer's Drug Discovery Foundation; Araclon Biotech; BioClinica, Inc. ; Biogen; Bristol-Myers Squibb Company; CereSpir, Inc.; Cogstate; Eisai Inc.; Elan Pharmaceuticals, Inc.; Eli Lilly and Company; EuroImmun; F. Hoffmann-La Roche Ltd and its affiliated company Genentech, Inc.; Fujirebio; GE Healthcare; IXICO Ltd.; Janssen Alzheimer Immunotherapy Research & Development, LLC.; Johnson & Johnson Pharmaceutical Research & Development LLC.; Lumosity; Lundbeck; Merck & Co., Inc.; Meso Scale Diagnostics, LLC.; NeuroRx Research; Neurotrack Technologies; Novartis Pharmaceuticals Corporation; Pfizer Inc.; Piramal Imaging; Servier; Takeda Pharmaceutical Company; and Transition Therapeutics. The Canadian Institutes of Health Research is providing funds to support ADNI clinical sites in Canada. Private sector contributions are facilitated by the Foundation for the National Institutes of Health (www.fnih.org). The grantee organization is the Northern California Institute for Research and Education, and the study is coordinated by the Alzheimer's Therapeutic Research Institute at the University of Southern California. ADNI data are disseminated by the Laboratory for Neuro Imaging at the University of Southern California.

Author Contributions

L.C. and M.G. conceptualized and designed the study. L.C., V.N., R.K., and S.H. acquired and analyzed the data. L.C., V.N., R.K., S.H., and M.G. drafted the text and prepared figures.

Potential Conflicts of Interest

Nothing to report.

References

- Minoshima S, Giordani B, Berent S, et al. Metabolic reduction in the posterior cingulate cortex in very early Alzheimer's disease. Ann Neurol 1997;42:85–94.
- de Leon MJ, Convit A, Wolf OT, et al. Prediction of cognitive decline in normal elderly subjects with 2-[(18)F]fluoro-2-deoxy-Dglucose/poitron-emission tomography (FDG/PET). Proc Natl Acad Sci U S A 2001;98:10966–10971.
- Chételat G, Desgranges B, de la Sayette V, et al. Mild cognitive impairment: can FDG-PET predict who is to rapidly convert to Alzheimer's disease? Neurology 2003;60:1374–1377.
- Fransson P, Marrelec G. The precuneus/posterior cingulate cortex plays a pivotal role in the default mode network: evidence from a partial correlation network analysis. Neuroimage 2008;42:1178–1184.
- Leech R, Sharp DJ. The role of the posterior cingulate cortex in cognition and disease. Brain 2014;137:12–32.
- Buckner RL, Andrews-Hanna JR, Schacter DL. The brain's default network: anatomy, function, and relevance to disease. Ann N Y Acad Sci 2008;1124:1–38.
- Greicius MD, Srivastava G, Reiss AL, Menon V. Default-mode network activity distinguishes Alzheimer's disease from healthy aging: evidence from functional MRI. Proc Natl Acad Sci U S A 2004;101:4637–4642.
- Cruchaga C, Kauwe JSK, Harari O, et al. GWAS of cerebrospinal fluid tau levels identifies risk variants for Alzheimer's disease. Neuron 2013;78:256–268.
- Ramanan VK, Risacher SL, Nho K, et al. GWAS of longitudinal amyloid accumulation on 18F-florbetapir PET in Alzheimer's disease implicates microglial activation gene IL1RAP. Brain 2015;138:3076–3088.
- Engler H, Forsberg A, Almkvist O, et al. Two-year follow-up of amyloid deposition in patients with Alzheimer's disease. Brain 2006;129:2856–2866.
- Kadir A, Almkvist O, Forsberg A, et al. Dynamic changes in PET amyloid and FDG imaging at different stages of Alzheimer's disease. Neurobiol Aging 2012;33:198.e1–14.
- Sabio G, Das M, Mora A, et al. A stress signaling pathway in adipose tissue regulated hepatic insulin resistance. Science 2008;322: 1539–1543.
- Westwood AJ, Beiser A, DeCarli C, et al. Insulin-like growth factor-1 and risk of Alzheimer dementia and brain atrophy. Neurology 2014;82:1613–1619.
- Weintraub S, Salmon D, Mercaldo N, et al. The Alzheimer's Disease Centers' Data Set (UDS): The neuropsychological test battery. Alzheimer Dis Assoc Disord 2009;23:91–101.
- Morris JC, Weintraub S, Chui HC, et al. The Uniform Data Set (UDS): Clinical and Cognitive Variables and Descriptive Data From Alzheimer Disease Centers. Alzheimer Dis Assoc Disord 2006;20:210–216.
- Beekly DL, Ramos EM, Lee WW, et al. The National Alzheimer's Coordinating Center (NACC) database: the Uniform Data Set. Alzheimer Dis Assoc Disord 2007;21:249–258.
- Allen M, Carrasquillo MM, Funk C, et al. Human whole genome genotype and transcriptome data for Alzheimer's and other neurodegenerative diseases. Sci Data 2016;3:160089.

- Bennett DA, Schneider JA, Buchman AS, et al. Overview and findings from the rush Memory and Aging Project. Curr Alzheimer Res 2012;9:646–663.
- Bennett DA, Schneider JA, Arvanitakis Z, et al. Overview and findings from the religious orders study. Curr Alzheimer Res 2012;9: 628–645.
- Jagust WJ, Bandy D, Chen K, et al; Alzheimer's Disease Neuroimaging Initiative. The Alzheimer's Disease Neuroimaging Initiative positron emission tomography core. Alzheimers Dement 2010;6: 221–229.
- Saykin AJ, Shen L, Foroud TM, et al; Alzheimer's Disease Neuroimaging Initiative. Alzheimer's Disease Neuroimaging Initiative biomarkers as quantitative phenotypes: genetics core aims, progress, and plans. Alzheimers Dement 2010;6:265–273.
- Price AL, Patterson NJ, Plenge RM, et al. Principal components analysis corrects for stratification in genome-wide association studies. Nat Genet 2006;38:904–909.
- Auton A, Abecasis GR, Altshuler DM, et al. A global reference for human genetic variation. Nature 2015;526:68–74.
- Cingolani P, Platts A, Wang LL, et al. A program for annotating and predicting the effects of single nucleotide polymorphisms, SnpEff: SNPs in the genome of Drosophila melanogaster strain w 1118; iso-2; iso-3. Fly 2012;6:80–92.
- Chang CC, Chow CC, Tellier LC, et al. Second-generation PLINK: rising to the challenge of larger and richer datasets. Gigascience 2015;4:7.
- 26. Risch N, Merikangas K. The future of genetic studies of complex human diseases. Science 1996;273:1516–1517.
- Fine JP, Gray RJ, Fine JP, Gray RJ. A proportional hazards model for the subdistribution of a competing risk. J Am Stat Assoc 1999; 94:496–509.
- Scrucca L, Santucci A, Aversa F. Regression modeling of competing risk using R: an in depth guide for clinicians. Bone Marrow Transplant 2010;45:1388–1395.
- Westra H-J, Peters MJ, Esko T, et al. Systematic identification of trans eQTLs as putative drivers of known disease associations. Nat Genet 2013;45:1238–1243.
- Jansen R, Hottenga JJ, Nivard MG, et al. Conditional eQTL analysis reveals allelic heterogeneity of gene expression. Hum Mol Genet 2017;26:1444–1451.

- Potkin SG, Guffanti G, Lakatos A, et al; Alzheimer's Disease Neuroimaging Initiative. Hippocampal atrophy as a quantitative trait in a genome-wide association study identifying novel susceptibility genes for Alzheimer's disease. PLoS One 2009;4:e6501.
- Stage E, Duran T, Risacher SL, et al. The effect of the top 20 Alzheimer disease risk genes on gray-matter density and FDG PET brain metabolism. Alzheimers Dement (Amst) 2016;5:53–66.
- Yoon YS, Lee MW, Ryu D, et al. Suppressor of MEK null (SMEK)/protein phosphatase 4 catalytic subunit (PP4C) is a key regulator of hepatic gluconeogenesis. Proc Natl Acad Sci U S A 2010;107:17704–1779.
- Dupuis J, Langenberg C, Prokopenko I, et al. New genetic loci implicated in fasting glucose homeostasis and their impact on type 2 diabetes risk. Nat Genet 2010;42:105–116.
- Craft S. Insulin resistance syndrome and Alzheimer's disease: ageand obesity-related effects on memory, amyloid, and inflammation. Neurobiol Aging 2005;26(suppl 1):65–69.
- Baker LD, Cross DJ, Minoshima S, et al. Insulin resistance and Alzheimer-like reductions in regional cerebral glucose metabolism for cognitively normal adults with prediabetes or early type 2 diabetes. Arch Neurol 2011;68:51–57.
- Mosconi L, Tsui WH, Herholz K, et al. Multicenter standardized 18F-FDG PET diagnosis of mild cognitive impairment, Alzheimer's disease, and other dementias. J Nucl Med 2008;49:390–398.
- Wolff S, Ma H, Burch D, et al. SMK-1, an essential regulator of DAF-16-mediated longevity. Cell 2006;124:1039–1053.
- Libina N, Berman JR, Kenyon C. Tissue-specific activities of C. elegans DAF-16 in the regulation of lifespan. Cell 2003;115:489–502.
- Cohen E, Dillin A. The insulin paradox: aging, proteotoxicity and neurodegeneration. Nat Rev Neurosci 2008;9:759–767.
- De Felice FG, Vieira MN, Bomfim TR, et al. Protection of synapses against Alzheimer's-linked toxins: insulin signaling prevents the pathogenic binding of Abeta oligomers. Proc Natl Acad Sci U S A 2009;106:1971–1976.
- Carro E, Trejo JL, Gerber A, et al. Therapeutic actions of insulinlike growth factor I on APP/PS2 mice with severe brain amyloidosis. Neurobiol Aging 2006;27:1250–1257.
- Herold C, Hooli B V, Mullin K, et al. Family-based association analyses of imputed genotypes reveal genome-wide significant association of Alzheimer's disease with OSBPL6, PTPRG, and PDCL3. Mol Psychiatry 2016;21:1608–1612.